

## BROAD REPERTOIRE OF T CELL AUTOREACTIVITY DIRECTLY FROM ISLETS OF DONORS WITH TYPE 1 DIABETES (T1D)

Jenny Aurielle B. Babon<sup>1</sup>, Megan E. DeNicola<sup>1</sup>, David M. Blodgett<sup>1</sup>, Inne Crèvecoeur<sup>2</sup>, Thomas S. Buttrick<sup>3</sup>, René Maehr<sup>4</sup>, Rita Bottino<sup>5,6</sup>, Ali Naji<sup>7</sup>, John Kaddis<sup>8</sup>, Wassim Elyaman<sup>3</sup>, Eddie A. James<sup>9</sup>, Rachana Haliyur<sup>10</sup>, Marcela Brissova<sup>10</sup>, Lut Overbergh<sup>2</sup>, Chantal Mathieu<sup>2</sup>, Thomas Delong<sup>11</sup>, Kathryn Haskins<sup>11</sup>, Alberto Pugliese<sup>12</sup>, Martha Campbell-Thompson<sup>13</sup>, Clayton Mathews<sup>13</sup>, Mark A. Atkinson<sup>13</sup>, Alvin C. Powers<sup>10,14,15</sup>, David M. Harlan<sup>1</sup>, Sally C. Kent<sup>1</sup>

<sup>1</sup>Division of Diabetes, Diabetes Center of Excellence, Department of Medicine, University of Massachusetts Medical School; <sup>2</sup>Laboratory for Clinical and Experimental Endocrinology, Department of Clinical and Experimental Medicine, KU Leuven, Leuven, Belgium; <sup>3</sup>Ann Romney Center for Neurologic Diseases, Brigham and Women's Hospital and Harvard Medical School, Boston, MA; <sup>4</sup>Program in Molecular Medicine, Diabetes Center of Excellence, University of Massachusetts Medical School; <sup>5</sup>Institute of Cellular Therapeutics, Allegheny-Singer Research Institute, Pittsburgh, PA; <sup>6</sup>Department of Biological Sciences, Carnegie Mellon University, Pittsburgh, PA; <sup>7</sup>Institute for Diabetes, Obesity, and Metabolism, University of Pennsylvania School of Medicine, Philadelphia, PA; <sup>8</sup>Department of Information Sciences, Beckman Research Institute, City of Hope, Duarte, CA; <sup>9</sup>Benaroya Research Institute at Virginia Mason, Seattle, WA; <sup>10</sup>Division of Diabetes, Endocrinology and Metabolism, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN; <sup>11</sup>Department of Immunology and Microbiology, University of Colorado School of Medicine, Denver, Anschutz Medical Campus, Aurora, CO; <sup>12</sup>Diabetes Research Institute, University of Miami, Miami, FL; <sup>13</sup>Departments of Pathology, Immunology, and Laboratory Medicine, University of Florida, Gainesville, FL; <sup>14</sup>Department of Molecular Physiology and Biophysics, Vanderbilt University, Nashville, TN; <sup>15</sup>VA Tennessee Valley Healthcare System, Nashville, TN

Type 1 diabetes (T1D) is an autoimmune disease characterized by the infiltration of lymphocytes into the insulin-producing  $\beta$ -cells in the pancreas. We have isolated live T cells sorted or grown directly from the isolated, handpicked islets of human donors with T1D. We received ~500 islet equivalent EQ of variable purity (10-90%) from 12 donors with T1D (disease duration 0.42-20 years) and from seven control donors and two donors with type 2 diabetes (T2D). A total of 321 T cell lines and clones were derived from the islets of donors with T1D (3 lines from the 9 control donors). These are 131 CD4<sup>+</sup> lines and clones, 47 CD8<sup>+</sup> lines and 143 lines that contain both CD4<sup>+</sup> and CD8<sup>+</sup> T cells. From 50 lines and clones examined to date, we have determined the autoreactivity of 19 and have seen a broad repertoire of T cell autoreactivity in the islets, including characterized targets and post-translationally modified targets. Autoreactivity of CD4<sup>+</sup> T cell lines was to three different peptides from glutamic acid decarboxylase 65 (GAD; GAD<sub>115-127</sub>, GAD<sub>274-286</sub>, GAD<sub>555-567</sub>), proinsulin<sub>76-90</sub>, and to chromogranin A or proinsulin expressed by DR4+DQ8+ B cells transduced with lentivirus containing constructs with the open reading frames corresponding to whole autoantigens. Reactivity to modified peptides included the glucose-regulated protein 78 and islet amyloid polypeptide with arginine to citrulline modifications (GRP78<sub>292-305</sub>(Arg-Cit<sub>297</sub>) and IAPP<sub>65-84</sub>(Arg-Cit<sub>73, 81</sub>)), deaminations (IA-2<sub>545-562</sub>(Gln-Glu<sub>548, 551, 556</sub>)), and to several insulin hybrid peptides. These autoreactive CD4<sup>+</sup> T cell lines and clones secreted only pro-inflammatory cytokines (IFN- $\gamma$ , TNF $\alpha$ ) upon peptide stimulation. For CD8<sup>+</sup> T cells from islets, from one donor with T1D, we saw binding of a pool of HLA-A2 pentamers loaded with insulin B<sub>10-18</sub>, IA-2<sub>797-805</sub> and insulin specific glucose-6-phosphatase catalytic subunit related protein, IGRP<sub>265-273</sub>. These results have implications for the development of successful prevention and reversal therapeutic strategies in T1D.

### **Contact:**

Jenny Aurielle B. Babon, Ph.D.  
University of Massachusetts Medical School  
[Jenny.babon@umassmed.edu](mailto:Jenny.babon@umassmed.edu)