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# Temporal Regulation of Nuclear Factor One Occupancy by Calcineurin/NFAT Governs a Voltage-Sensitive Developmental Switch in Late Maturing Neurons

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Dendrite and synapse development are critical for establishing appropriate neuronal circuits, and disrupted timing of these events can alter neural connectivity. Using microarrays, we have identified a nuclear factor I (NFI)-regulated temporal switch program linked to dendrite formation in developing mouse cerebellar granule neurons (CGNs). NFI function was required for upregulation of many synapse-related genes as well as downregulation of genes expressed in immature CGNs. Chromatin immunoprecipitation analysis revealed that a central feature of this program was temporally regulated NFI occupancy of late-expressed gene promoters. Developing CGNs undergo a hyperpolarizing shift in membrane potential, and depolarization inhibits their dendritic and synaptic maturation via activation of calcineurin (CaN) (Okazawa et al., 2009). Maintaining immature CGNs in a depolarized state blocked NFI temporal occupancy of late-expressed genes and the NFI switch program via activation of the CaN/nuclear factor of activated T-cells, cytoplasmic (NFATc) pathway and promotion of late-gene occupancy by NFATc4, and these mechanisms inhibited dendritogenesis. Conversely, inhibition of the CaN/NFATc pathway in CGNs maturing under physiological nondepolarizing conditions upregulated the NFI switch program, NFI temporal occupancy, and dendrite formation. NFATc4 occupied the promoters of late-expressed NFI program genes in immature mouse cerebellum, and its binding was temporally downregulated with development. Further, NFI temporal binding and switch gene expression were upregulated in the developing cerebellum of *Nfatc4* (–/–) mice. These findings define a novel NFI switch and temporal occupancy program that forms a critical link between membrane potential/CaN and dendritic maturation in CGNs. CaN inhibits the program and NFI occupancy in immature CGNs by promoting NFATc4 binding to late-expressed genes. As maturing CGNs become more hyperpolarized, NFATc4 binding declines leading to onset of NFI temporal binding and the NFI switch program.

## Introduction

The timing of dendrite formation and synaptogenesis is intimately involved in establishing unique patterns of neuronal connectivity (Deguchi et al., 2011; Tripodi and Arber, 2012). Accordingly, gene expression must be precisely coordinated in time to ensure that synaptic circuits properly form (Hippenmeyer et al., 2005; Di Cristo et al., 2007; Petrovic and

Hummel, 2008). Temporal dysregulation leading to altered synapse formation has been implicated in several neurodevelopmental disorders (Geschwind and Levitt, 2007; Leonardo and Hen, 2008; Meredith et al., 2012).

Cerebellar granule neurons (CGNs) function in cerebellar information processing via synaptic connections with mossy fiber inputs and Purkinje cell outputs (D'Angelo et al., 2011). CGNs have been studied extensively to elucidate mechanisms governing neuronal development, including dendritogenesis and synapse formation (Goldowitz and Hamre, 1998; Hall et al., 2000; Ito and Takeichi, 2009). CGN progenitors (CGNPs) proliferate in the external germinal layer (EGL) and generate immature neurons within the premigratory zone (PMZ). CGN cell bodies later migrate inwardly until reaching the internal granule cell layer (IGL), where postmigratory CGNs form mature dendrites and synaptic connections with input neurons. Numerous genes are sequentially expressed during these different developmental stages (Goldowitz and Hamre, 1998; Furuichi et al., 2011). Much of this developmental program is recapitulated in CGN cultures (Ito

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The authors declare no competing financial interests.

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and Takeichi, 2009; de la Torre-Ubieta et al., 2010) wherein CGNs and immature CGNs isolated from the EGL/PMZ (Raetzman and Siegel, 1999) differentiate into IGL-like cells upon plating (Manzini et al., 2006; Wang et al., 2011).

Nuclear factor I (NFI) transcription factors are important regulators of CGN maturation (Kilpatrick et al., 2012). They are expressed throughout CGN postmitotic development and regulate parallel fiber extension, migration, dendritogenesis, and synaptogenesis (Wang et al., 2007, 2010; Piper et al., 2011). Recently, NFI proteins were shown to control the dendritogenesis-linked expression of the  $\alpha 6$  GABA<sub>A</sub> receptor subunit (*Gabra6*) gene within the developing IGL (Wang et al., 2011). CGNs thus provide a robust model for exploring the programming of dendrite and synapse formation and the role of NFI proteins in these events (Kilpatrick et al., 2012).

CGNs undergo a shift in membrane potential as they differentiate, becoming increasingly hyperpolarized with maturation (Rossi et al., 1998; Cathala et al., 2003; Okazawa et al., 2009). This change in membrane potential has been implicated in the control of mouse CGN differentiation in culture and within the developing cerebellum (Nakanishi and Okazawa, 2006). In particular, depolarization of immature CGNs inhibits developmental changes in gene expression as well as dendritic and synaptic maturation via Ca<sup>2+</sup>-dependent activation of calcineurin (CaN) (Mellor et al., 1998; Ives et al., 2002; Sato et al., 2005; Iijima et al., 2009; Okazawa et al., 2009).

Here we describe an NFI-regulated temporal switch program that regulates late differentiation of postmigratory CGNs, including dendrite formation. A central feature of this program is delayed NFI occupancy of numerous genes that are temporally upregulated as CGNs mature. Further, we show that this NFI switch is directly linked to membrane potential-dependent mechanisms via its regulation by CaN activation of nuclear factor of activated T-cells, cytoplasmic (NFATc) transcription factors.

## Materials and Methods

**Animals and primary cultures.** Mouse CGNs were prepared from 6-d-old CD1 mouse pups of either sex as previously described (Wang et al., 2004). Individual culture experiments were performed using cells prepared from the same litter. Cells were plated at a density of  $5 \times 10^4$  cells/cm<sup>2</sup> onto chamber slides or cell culture dishes coated with poly-D-lysine/laminin (Invitrogen) in Neurobasal medium (Invitrogen) containing B-27 serum-free supplement (50 $\times$ ; Invitrogen) (Wang et al., 2004). The following reagents were added to culture medium  $\sim 1$  h after cell plating and were refreshed on 3 d *in vitro* (DIV): 5  $\mu$ M FK506 (Sigma), 1  $\mu$ M cyclosporin A (Sigma), 10  $\mu$ M nimodipine (Tocris Bioscience), 100  $\mu$ M kainic acid (KA; Sigma), vehicle control (dimethylsulfoxide). Cerebellar tissues were obtained from postnatal *Nfat4* (–/–) mice on a C57BL/6 background as well as from wild-type (WT) C57BL/6 mice (Charles River). *Nfia* (–/–) mice and WT littermates were on a C57BL/6NTac background.

**Plasmids and cell lines.** Self-inactivating lentiviruses expressing hemagglutinin (HA)-tagged NFI dominant repressor (NFI/EnR) or *Drosophila* engrailed repressor domain (EnR) were described previously (Wang et al., 2004). FLAG-tagged NFATc4 proteins [(constitutively active (NFATc4-Ala) and dominant-negative NFAT (dnNFAT)] were released from their expression vectors (Chow et al., 1999; Yang et al., 2002) and inserted into BamHI and XbaI sites of the lentiviral expression vector pHR<sup>+</sup>-cPPT-CMV-W-Sin18 (Wang et al., 2004). Human embryonic kidney 293T cells were grown in DMEM (Invitrogen) containing 10% heat-inactivated fetal bovine serum (Invitrogen).

**Lentivirus preparation and transduction of primary CGNs.** Lentiviruses were generated by cotransfection of 293T cells with lentiviral expression plasmid, psPAX2 and pCMV-VSVG, then concentrated and titered as previously described (Wang et al., 2005). CGN cultures were transduced on 0 DIV using MOI = 2.5–3, yielding 80–90% cell transduction.

**RNA isolation and quantitative real-time PCR.** RNA was extracted from tissues or cultured cells using Tri reagent (Sigma), processed with DNA-free kit (Ambion), and complementary DNAs were prepared as described previously (Wang et al., 2007). Real-time PCRs were performed in triplicate using a StepOnePlus Real-Time PCR System (Applied Biosystems). Target transcripts were normalized to 18S rRNA by the  $2^{-\Delta\Delta Ct}$  method, and differences relative to control samples were calculated using  $2^{-\Delta\Delta Ct}$  as previously described (Wang et al., 2011). Numerical values for  $2^{-\Delta\Delta Ct} < 1.0$  were recalculated as control relative to experimental data and expressed as negative numbers. Real-time PCR primer sequences are available upon request.

**Microarray and bioinformatic analyses.** Total RNA was purified from CGN cultures with an RNeasy Plus Micro Kit (Qiagen). RNA quality analysis and hybridizations to GeneChip Mouse Genome 430A,B or 430 2.0 arrays (Affymetrix) were performed by the Genomics Core Facility of the University of Massachusetts Medical School. Differentially expressed genes for temporal and NFI arrays were analyzed separately. For NFI arrays (four NFI/EnR expressed vs four EnR control biological replicates), gene expression values in the Affymetrix cel files were first background corrected and normalized together using RMA algorithm (Bioconductor R package). To identify genes with robust expression, we removed probes with low expression (i.e., expression value below 40th percentile in at least seven of eight arrays). For multiple probes mapping to the same gene, only the probe showing the highest expression change for each gene was retained. Finally, moderated *t* statistics and FDR correction (limma R package) were used to assess the significance of differential expression. NFI differentially regulated genes were defined as probes with FDR *p* value < 0.15 and fold change > 1.5. Identical steps for signal normalization, background noise correction, and statistical testing were applied to temporal arrays (1.5 DIV vs 6 DIV, three biological replicates each). Differentially expressed genes were defined as having probes with FDR *p* value < 0.1 and fold change > 2. We used more stringent criteria to define temporally regulated genes since the variability between replicates appeared smaller. Greater variability among the NFI/EnR array data likely reflected differing infectivities of distinct lentiviral preparations used for independent experiments. Finally, NFI-regulated and temporally regulated overlap genes showed significant differential expression for the same probes in both NFI and temporal datasets. Chi-square tests were used to assess the significance of the overall overlap dataset as well as its partitioning into four NFI/temporal categories based on the directionality of expression changes.

Citations related to synaptic function, signaling, and excitability were identified in the PubMed database. Mouse gene promoter sequences were extracted from GenBank and were scanned for NFI consensus sites using the TRANSFAC database.

**Chromatin immunoprecipitation assays.** Chromatin immunoprecipitation (ChIP) assays were performed as previously described (Wang et al., 2011). For tissues, nuclei were first purified by Percoll gradient centrifugation (Wang et al., 2011) or by differential centrifugation (1400  $\times$  g for 3 min, 4°C) in 0.25 M sucrose. Samples were assayed by PCR using polyacrylamide gels or real-time PCR. PCR data were expressed as a percentage of input using the  $2^{-\Delta\Delta Ct}$  method. ChIP antibodies were to *Xenopus* NFIB1 (xNFIB1) (Puzianowska-Kuznicka and Shi, 1996), NF1A (Active Motif; 39036), NFATc4 (Santa Cruz Biotechnology; sc-1153), and FLAG epitope (Sigma; F1804). Pre-immune serum or normal rabbit IgG (Millipore; PP64) were used as negative controls. PCR primer sequences for ChIP assays are available upon request.

**Immunofluorescence and dendritogenesis assay.** Immunofluorescence was performed as in previous studies (Wang et al., 2007). Briefly, primary CGNs were cultured in coated chamber slides and then were fixed on 1.5 DIV or 2 DIV with 4% paraformaldehyde/PBS. Cells were permeabilized with 1% Triton X solution, incubated with 5% normal goat serum followed by primary antibodies at 4°C overnight and then Cy3-conjugated goat anti-rabbit IgG (NG1807775; Millipore). Dendrite assays used anti-Map2 antibody (Millipore; AB5622) and 1  $\mu$ g/ml Bisbenzimidazole (Sigma) to stain nuclei. Dendrite length was measured with Image Pro Plus 6.0 software. Lentiviral titrations used antibodies for HA (NFI/EnR and EnR; C29F4, Cell Signaling Technology) or FLAG (NFATc4 proteins; M2 antibody, Sigma).

**Western blot.** Total nuclear proteins were prepared from cultured CGNs as previously described (Wang et al., 2004). Purified NFIA, NFIB, and NFIX proteins were purchased from Abnova. Protein samples were separated on 8% SDS-polyacrylamide gels and transferred onto Pure Nitrocellulose membranes (GE Water & Process Technologies). Following blocking for 1 h, blots were incubated with primary antibodies at 4°C overnight and then with horseradish peroxidase-conjugated secondary antibody at room temperature for 1 h. Bound antibodies were detected with a chemiluminescent substrate (Thermo Scientific). Primary antibodies used were as follows: xNFIB antibody (1:200), pan-NFI antibody (1:1000; Santa Cruz Biotechnology, sc-5567), and anti-Histone H3 (1:1000; Abnova, PAB0653).

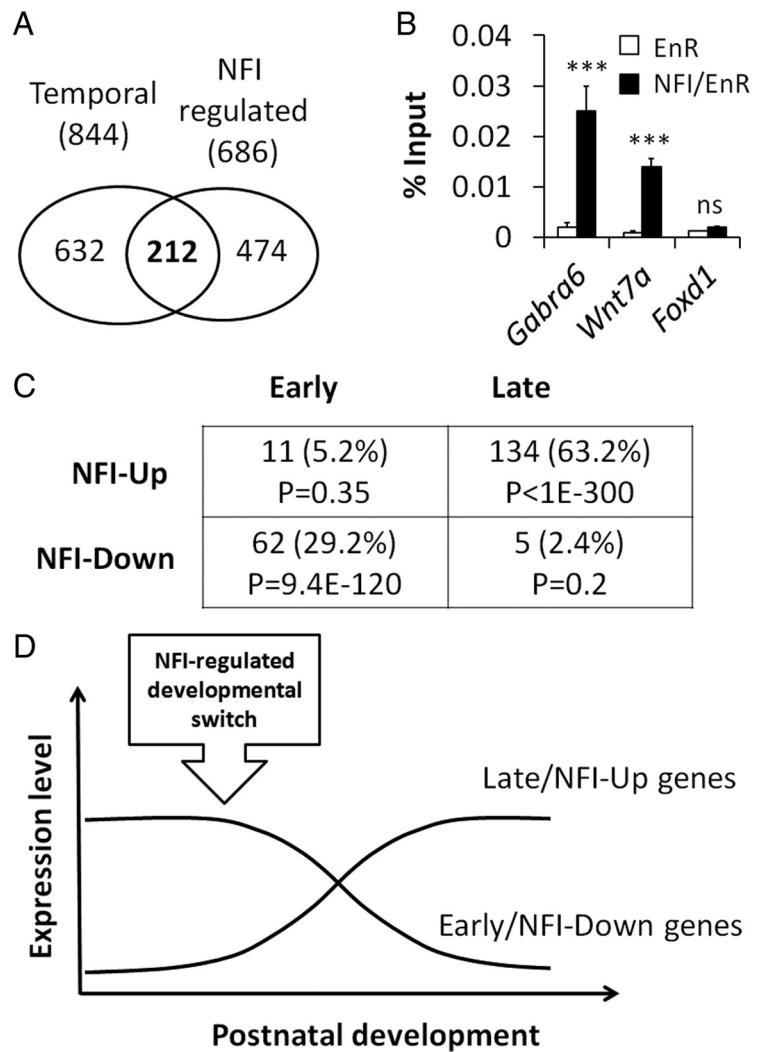
**NFAT transactivation and CaN-dependent dephosphorylation assays.** To determine NFAT transcriptional activity, cultured CGNs were transduced with an NFAT dual luciferase reporter lentiviral vector (Na et al., 2010) (kindly provided by the van den Brink lab, Memorial Sloan-Kettering Cancer Center, NY). NFAT trans-activation was quantified by measuring NFAT-inducible click-beetle red luciferase (CBRLuc) and normalizing activity to a constitutively expressed membrane-anchored *Gussia* luciferase (extGLuc) using the Dual-Glo Luciferase Assay System (Promega). CaN-dependent dephosphorylation activity was measured using a cellular assay kit (Enzo Life Sciences) according to the manufacturer's instructions. Data were normalized to protein concentration.

**Statistics.** In all quantitative studies other than microarrays, data were statistically analyzed using the two-tail *t* test. Results were expressed as the mean  $\pm$  SD, and *p* values < 0.05 were considered significant.

## Results

### NFI controls a developmental switch in maturing CGNs

NFI proteins regulate several developmentally expressed genes in maturing CGNs (Wang et al., 2007, 2010). Here we determined whether these proteins controlled a larger temporal program using a 2D microarray analysis of temporally regulated and NFI-regulated genes. For temporal genes, we compared immature CGNs (1.5 DIV) with more differentiated (6 DIV) cultures. CGN cultures (1.5 DIV) are largely postmitotic (Wang et al., 2005) and exhibit a morphology and pattern of gene expression consistent with immature CGNs *in vivo*, while 6 DIV cultures resemble IGL-like CGNs (Raetzman and Siegel, 1999; Manzini et al., 2006; Wang et al., 2010, 2011). This analysis identified 844 Temporal-Up or Temporal-Down genes (Fig. 1A). NFI-regulated genes were assayed in more mature 6 DIV CGN cultures that were transduced at the time of plating (0 DIV) with NFI dominant repressor (HA-NFI/EnR) or control (HA-EnR) proteins using lentiviruses (Wang et al., 2004). This identified 686 NFI-regulated genes that were either suppressed or elevated by NFI/EnR (Fig. 1A). Based on its dominant repressor function, NFI/EnR should inhibit expression of genes directly activated by NFI proteins. Consistent with this, HA ChIP detected NFI/EnR bound to the *Gabra6* and



**Figure 1.** Identification of an NFI developmental switch program in developing CGNs. **A**, Diagram showing temporal (844 genes) and NFI-regulated (686 genes) microarray datasets and their overlap (212 genes). **B**, ChIP pPCR assays were performed for HA-tagged protein binding to NFI sites within the *Gabra6* and *Wnt7a* promoters using chromatin from 6 DIV CGN cultures transduced with NFI/EnR and EnR proteins on 0 DIV. *Foxd1*/brain factor 2 coding sequences served as negative control genomic sequences. Negative control (IgG) ChIP signal was subtracted in each case.  $***p < 0.001$ ; ns, no significant difference. Similar results were obtained in two independent experiments. **C**, Segregation of NFI/Temporal overlap genes into different subsets based on direction of regulation. Numbers of genes, percentages of total overlap genes, and statistical significance (*p* values) are indicated for each subset. **D**, Model for the NFI-regulated switch program in developing CGNs. As CGNs mature, NFI proteins upregulate Late genes and downregulate Early genes.

*Wnt7a* NFI sites (Fig. 1B). Genes upregulated by the dominant repressor may have been indirectly derepressed via suppression of intermediary regulators required for their downregulation. Other mechanisms also are possible; e.g., certain genes may have been directly repressed by endogenous NFI proteins, but less efficiently by the dominant repressor resulting in their upregulation. The latter question was not further examined here.

Overlap analysis identified 212 genes that were both NFI and temporally regulated (Fig. 1A). This overlap was highly significant ( $p = 4.29 \times 10^{-22}$ ). Temporal genes were enriched  $\sim 8$ -fold within the NFI-regulated set relative to non-NFI regulated genes, indicating the importance of NFI proteins in developmental gene regulation. Overlap genes segregated into four gene subsets based on their temporal patterns and regulation by endogenous NFI proteins (Fig. 1C): Early/NFI-Down, Early/NFI-Up, Late/NFI-Down, Late/NFI-Up. Early and Late genes were temporally downregulated or upregulated on 6 DIV; NFI-Up genes were

**Table 1. Validation of NFI switch gene regulation in culture and *in vivo* using qRT-PCR**

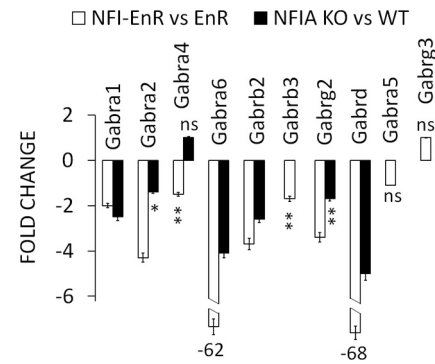
Gene symbol	Gene ID	6 DIV versus 0 DIV	P22 versus P7	NFI-EnR versus EnR	<i>Nfia</i> <sup>-/-</sup> versus <i>Wt</i>
<b>Late/NFI-Up</b>					
<i>Wnt7a</i>	22421	12	9.9	-17	-2.9
<i>Nptx1</i>	18164	35	44	-71	-3.2
<i>Rps6ka1</i>	20111	39	7.1	-42	-3.5
<i>Nab2</i>	17937	13	8.2	-10	-3.1
<i>Tesc</i>	57816	142	43	-7.8	-6.8
<i>Camkk2</i>	207565	12	8.1	-2.3**	-3.8
<i>Camk4</i>	12326	23	17	-22	-5.5
<i>Egr1</i>	13653	-13	4.6	-6.4	1.1 ns
<i>Ets2</i>	23872	—	—	-3.1	-7.3
<b>Early/NFI-Down</b>					
<i>Dcx</i>	13193	-5.8	-142	2.6**	2.6**
<i>Neurod6</i>	11922	-3.4	-11	1.7*	1.4*
<i>Tgfb2</i>	21808	-3.4	-4.5	1.5*	1.4*
<i>Met</i>	17295	-1.3*	-4	6.7	-1.6*
<i>Ect2</i>	13605	-20	-88	2.8	3.6
<i>Nhlh1</i>	18071	—	—	—	2.3
<b>Late/NFI-Down</b>					
<i>Pcdh17</i>	219228	2.7**	-3.6	14	-2.7**
<b>Early/NFI-Up</b>					
<i>Nrcam</i>	319504	3.9	-1.6*	-2.1*	-2.2**
<i>Cdh10</i>	320873	2.5**	-2.9**	-6.4	-1.2 ns
<b>Additional genes</b>					
<b>Late/NFI-Up</b>					
<i>Kcnab1</i>	16497	—	—	-4.8	—
<i>Slc12a5</i>	57138	—	—	-2.3**	—
<i>Kcnk3</i>	16527	—	—	-6	—
<i>Syn2</i>	20965	—	—	-2.3**	—
<b>Nonoverlap</b>					
<i>Id2</i>	15902	—	—	6.4	—
<i>Scrg1</i>	20284	—	—	-5.6	—
<i>Hopx</i>	74318	—	—	-5.3	—
<i>Tiam1</i>	21844	—	—	-20.0*	-3.7

Data are expressed as the relative difference in expression; negative values indicate reduced expression relative to controls. Dashes, not determined.  $p < 0.001$  except \* $p < 0.05$ , \*\* $p < 0.01$ ; ns, no significant difference. Data are representative of three independent experiments.

upregulated by NFI (repressed by NFI/EnR) while NFI-Down genes were inhibited by NFI (elevated by NFI/EnR) in 6 DIV cultures.

Nearly all (92%) NFI temporal genes clustered into two groups: Early/NFI-Down (62 genes; 29% of overlap genes) and a predominant set of Late/NFI-Up genes (134; 63% of total) (Fig. 1C). This skewed distribution was also highly statistically significant (Fig. 1C), and it suggested a “developmental switch” mechanism in which NFI proteins downregulate genes expressed in immature CGNs and upregulate late-expressed genes (Fig. 1D). Based on this model, dominant repression of NFI function “trapped” CGNs in an immature state, stimulating or preventing the temporal downregulation of Early/NFI-Down gene expression, and directly or indirectly suppressing expression of Late/NFI-Up genes.

Regulation by NFI/EnR was confirmed in CGN cultures for a total of 34 genes using qRT-PCR (Table 1; Fig. 2), including several transcripts that did not reach significance in the overlap set but were of regulatory or functional interest. For 25 genes we also determined NFI regulation *in vivo* using P15 *Nfia*-null mouse cerebellum, which showed identical trends with NFI/EnR data with the exception of *Egr1* (Table 1). This may reflect a compensatory or required role for other NFI family members in



**Figure 2.** NFI controls GABA<sub>A</sub> receptor switch gene expression. qRT-PCR assays of GABA<sub>A</sub> receptor subunit genes that undergo developmental changes in gene expression during CGN maturation. Changes were assessed in 6 DIV CGN cultures transduced on 0 DIV with NFI/EnR or EnR lentivirus, and in P15 *Nfia* (+/+) (WT) and (-/-) (KO) cerebella. Larger fold changes for the *Gabra6* and *Gabrd* genes are shown numerically below the bars.  $p < 0.001$  except \* $p < 0.05$ , \*\* $p < 0.01$ ; ns, no significant difference. Data are representative of three independent experiments.

*Egr1* gene regulation. Temporal expression was also determined in culture and *in vivo* for 16 genes (Table 1), and 8 others (*Tiam1*, *Ets2*, *Kcnk3*, *Slc12a5*, and GABA<sub>A</sub> receptor genes (Fig. 2) were previously reported (Laurie et al., 1992; Sato et al., 2005). This revealed that the NFI switch was activated between P7 and P21, when postmigratory CGNs within the IGL form dendrites and synapses. We also identified several temporally regulated non-overlap genes that were not dependent on NFI (*Tle1*, *Scn2a1*, *Kcna1*, *Kcnk9*, *Prickle1*, *Rbfox3*) (data not shown).

### NFI controls GABA<sub>A</sub> receptor maturation and synapse-related gene expression

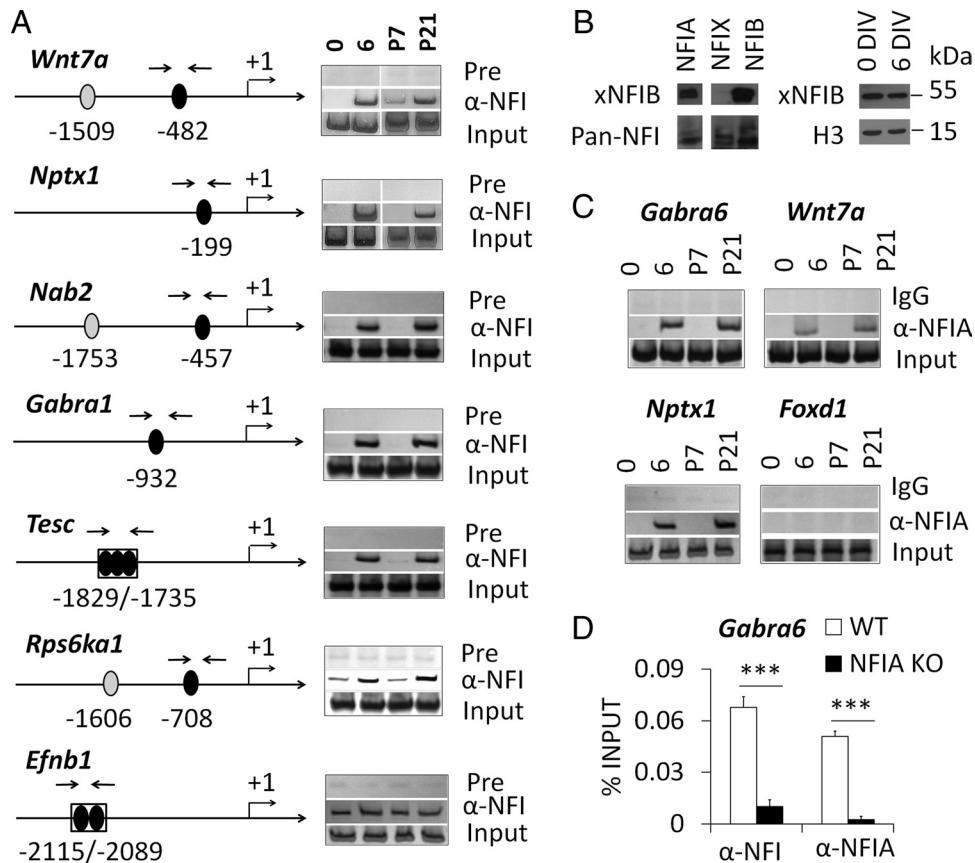
GABA<sub>A</sub> receptors undergo a developmental switch in subunit composition as postmigratory CGNs mature within the IGL, with  $\alpha 1$ -,  $\alpha 6$ -,  $\beta 2$ -,  $\beta 3$ -,  $\gamma 2$ -, and  $\delta$ -subunits being upregulated (Laurie et al., 1992; Zheng et al., 1993). This receptor switch is critical for controlling CGN excitability *in vivo* via GABA-induced tonic inhibition (Rossi and Hamann, 1998). *Gabra6* ( $\alpha 6$ ), *Gabra1* ( $\alpha 1$ ), *Gabrb2* ( $\beta 2$ ), and *Gabrg2* ( $\gamma 2$ ) were identified as Late/NFI-Up transcripts by microarrays, suggesting that NFI proteins regulated the GABA<sub>A</sub> receptor switch in CGNs. qRT-PCR confirmed that expression of these four genes was reduced by NFI dominant repression in CGN cultures and by *Nfia* deficiency *in vivo* (Fig. 2). Further, expression of the GABA<sub>A</sub> switch genes *Gabrb3* ( $\beta 3$ ) and *Gabrd* ( $\delta$ ) also were dependent on NFI. Interestingly, the *Gabra2* ( $\alpha 2$ ) and *Gabra4* ( $\alpha 4$ ) genes, which are transiently upregulated in maturing CGNs (Laurie et al., 1992), also were regulated by NFI proteins (Fig. 2), although *Gabra4* was only modestly regulated. In contrast, the developmentally unregulated *Gabra5* ( $\alpha 5$ ) and downregulated *Gabrg3* ( $\gamma 3$ ) receptor genes (Laurie et al., 1992) were insensitive to NFI functional disruption (Fig. 2).

Initial analysis of overlap genes showed enrichment of gene functions related to cell division in the Early/NFI-Down set (e.g., *Ccna2*, *Cks2*) and neuronal differentiation in Late/NFI-Up genes (e.g., *Ntrk3*, *Slit3*, *Wnt7a*), among others (data not shown). Since the NFI switch is activated as CGNs form synapses within the IGL, we performed a literature analysis of known associations of NFI-regulated temporal genes with dendritic and synaptic structure/function, excitability, and signaling. Remarkably, 63 Late/NFI-Up genes (47%) had known links to these events (Table 2), consistent with their temporal upregulation by NFI. In contrast,

**Table 2. NFI/Temporal overlap genes with citations related to regulation of synaptic/dendritic development, synaptic function, and signaling**

Late/Nfi-Up							Early/Nfi-Down
<i>Sorl1</i>	<i>Cbx6</i>	<i>Stxbp5l</i>	<i>Chn2</i>	<i>Pak1</i>	<i>Slc12a5</i>	<i>ApoE</i>	<i>Stmn1</i>
<i>Klf9</i>	<i>Zdhc18</i>	<i>Wnt7a</i>	<i>Dact1</i>	<i>Eps8</i>	<i>Rasgrf1</i>	<i>Cbln3</i>	<i>Cdc20</i>
<i>Gas6</i>	<i>Ncdn</i>	<i>Atp1a2</i>	<i>Adam11</i>	<i>Kcnk3</i>	<i>Iqsec1</i>	<i>Ntrk3</i>	<i>Nnat</i>
<i>Vsnl1</i>	<i>Col19a1</i>	<i>Trpm3</i>	<i>Gsg1l</i>	<i>Tyr03</i>	<i>Rap1gap</i>	<i>Bcan</i>	<i>Tgfb2</i>
<i>Lrrc4c</i>	<i>Tmeff2</i>	<i>Sybu</i>	<i>Cacna2d3</i>	<i>Slc4a4</i>	<i>Rora</i>	<i>Etv1</i>	<i>Slitrk5</i>
<i>Ptprr</i>	<i>Gas7</i>	<i>Syn2</i>	<i>Tem163</i>	<i>Nck1</i>	<i>Dlgap1</i>	<i>Gabra6*</i>	<i>Sncap</i>
<i>Dpp10</i>	<i>Ppfi4</i>	<i>Unc13c</i>	<i>Syt1</i>	<i>Lrrtm3</i>	<i>Inadl</i>	<i>Gabrb2*</i>	<i>Dpysl5</i>
<i>Nup210</i>	<i>Boc</i>	<i>Lrr3cb</i>	<i>Gprc5b</i>	<i>Sparc</i>	<i>Kcnab1</i>	<i>Gabrg2*</i>	
<i>Smarcd3</i>	<i>Gprc5b</i>	<i>Kcnh1</i>	<i>Sept8</i>	<i>Prune2</i>	<i>Nptx1</i>	<i>Gabra1*</i>	

Hyperlinks to publications relevant to synapse and dendrite function, localization, regulation, signaling, or excitability are provided in the online version of the article. \*See text.



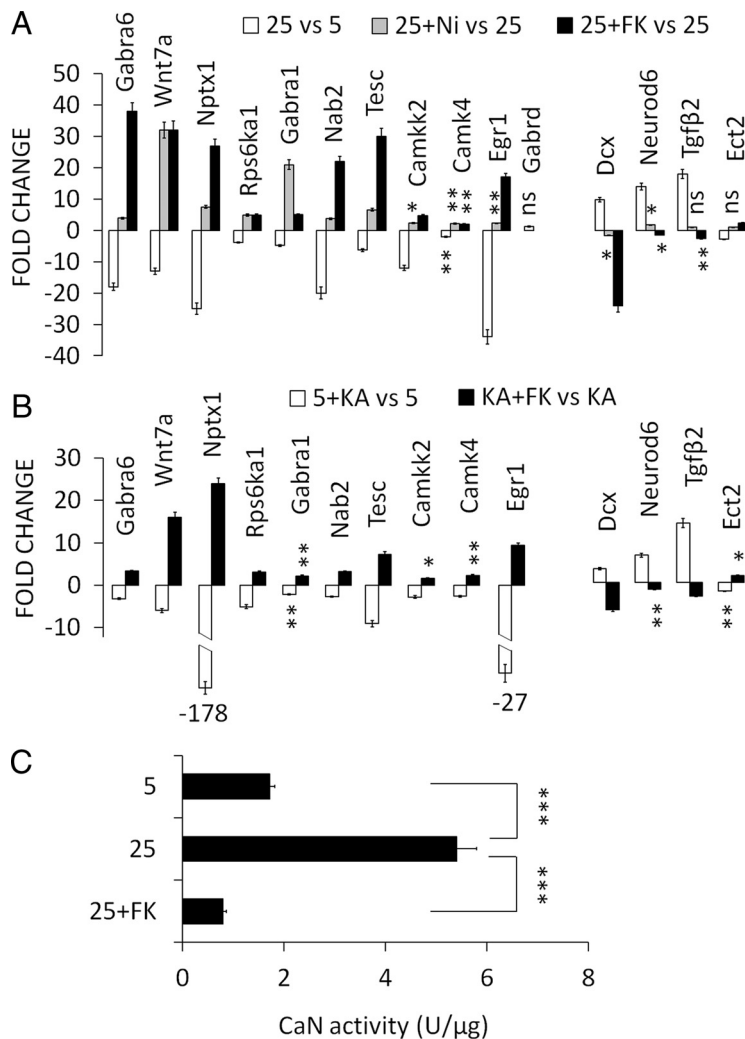
**Figure 3.** NFI occupancy of Late/NFI-Up genes is temporally regulated. **A**, Left, Schematic of predicted NFI binding sites in the 5'-flanking regions of the indicated gene promoters. Open ovals, weak NFI binding sites; filled ovals, strong NFI binding sites. Arrows, position of primers used for ChIP qPCR. Right, Temporal NFI ChIP results. Pre, pre-immune serum;  $\alpha$ -NFI, anti-xNFIB1 serum. Similar ChIP results were obtained in two or three experiments for each gene. **B**, Left, Characterization of the specificity of xNFIB1 antibody for expressed NFI family proteins on Western blots. Pan-NFI antibody staining of the same lanes is shown below. Right, NFI protein expression in nuclear extracts of immature and more differentiated CGN cultures detected with xNFIB1 antibody. Histone H3 (H3) served as a loading control. Data are representative of two independent experiments. **C**, ChIP assays showed that NFIA binds in a temporally delayed manner to Late/NFI-Up genes. *Foxd1* genomic sequences served as a negative control. Identical patterns were observed in two independent experiments. **D**, ChIP/qPCR assays of P15 *Nfia* (+/+) (WT) and (-/-) (KO) mouse cerebella confirmed that occupancy of the *Gabra6* NFI site as detected by xNFIB1 ( $\alpha$ -NFI) and NFIA antibodies was dependent on NFIA. \*\*\* $p < 0.001$ . Negative control antibody values were subtracted to yield specific antibody ChIP/qPCR signals. Data are representative of two independent experiments.

only seven Early/Down genes (11%) had known synapse-related activities or localization, indicating continuing functions of this small gene cohort in more mature CGNs following their developmental downregulation.

### NFI occupancy of numerous Late/NFI-Up genes is temporally regulated

The timing of *Gabra6* gene expression during CGN development is linked to delayed NFI binding to its promoter (Wang et al., 2011). Further, NFI proteins and their nuclear DNA binding activity are constitutively expressed during CGN maturation in cul-

ture and *in vivo* (Wang et al., 2011), indicating that NFI temporal occupancy of the *Gabra6* gene is regulated at the level of chromatin interactions. We therefore examined whether temporally regulated NFI chromatin occupancy was a more global mechanism for six other Late/NFI-Up genes (*Wnt7a*, *Nptx1*, *Nab2*, *Gabra1*, *Tesc*, *Rps6ka1*) using ChIP. The 5'-flanking regions for each gene contained one or more consensus NFI sites, at least one of which showed detectable NFI occupancy (Fig. 3A). In each case, the timing of NFI binding mirrored temporal expression: binding was undetectable or low in 0 DIV cells consisting of premigratory CGNPs/CGNs derived from the EGL/PMZ (Raetzman and



**Figure 4.** Regulation of the NFI switch program by membrane depolarization and Ca<sup>2+</sup>. qRT-PCR assays in **A** and **B** were performed for Late/NFI-Up genes (left) and Early/NFI-Down genes (right). **A**, Depolarization by 25 mM KCl culture medium (25) inhibits the NFI gene switch in 6 DIV CGNs, which is reversed by inhibitors of L-type VGCCs (nimodipine, Ni) and of Ca<sup>2+</sup> (FK506, FK). CGNs cultured in medium containing 5 mM KCl (5). **B**, KA inhibits NFI switch gene expression in CGNs cultured to 6 DIV in 5 mM KCl-containing medium (5), which is reversed by FK506. Large fold-change values for Nptx1 and Egr1 are shown numerically. *p* < 0.001 except \**p* < 0.05, \*\*\**p* < 0.01; ns, no significant difference. **C**, Ca<sup>2+</sup> activity is stimulated in depolarized CGN cultures. CGNs were cultured in medium containing either 5 mM KCl (5) or 25 mM KCl (25) ± FK506 to 6 DIV and Ca<sup>2+</sup> activity was determined in soluble cell extracts. Stimulation of phosphatase activity in depolarizing medium was prevented by the Ca<sup>2+</sup> inhibitor FK506. \*\*\**p* < 0.001. Data in **A–C** are representative of three independent experiments.

Siegel, 1999) and in the P7 cerebellum composed mainly of immature postmigratory CGNs from the IGL (Wang et al., 2011), but was markedly elevated in more mature (6 DIV) CGN cultures and P21 mouse cerebellum. In contrast, the *Efnb1* gene, which is a direct NFI target (Wang et al., 2007) expressed at similar levels in immature and more differentiated CGNs (data not shown), exhibited constitutive NFI binding to its cognate promoter site (Fig. 3A). Delayed NFI occupancy was thus a common and specific temporal mechanism for many Late/NFI-Up genes in CGNs.

Temporal ChIP assays were performed with an anti-*Xenopus* NFIB1 (-xNFIB1) antibody, which recognized mammalian NFIB and NFIA, but not NFIX (Fig. 3B). Further, the abundance and mobility of NFI proteins detected by this antibody did not vary significantly with CGN maturation (Fig. 3B), as shown previously using pan-NFI antibodies (Wang et al., 2011). Since knock-out studies implicated NFIA in regulation of the switch program, we also confirmed its temporal occupancy for several Late/Up

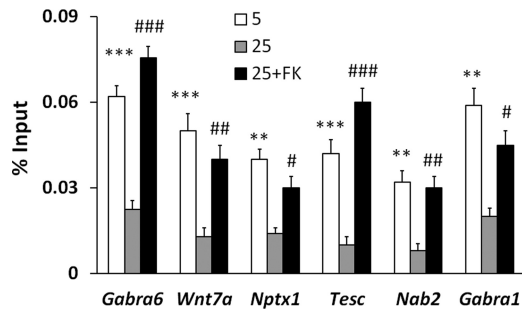
genes in developing CGNs and cerebellum (Fig. 3C). Further, we confirmed that ChIP signals for both anti-xNFIB1 and NFIA antibodies were markedly reduced in chromatin from P15 *Nfia*-null mouse cerebella (Fig. 3D). Thus, NFIA appears to be a direct regulator of the NFI temporal program in CGNs via its delayed occupancy of Late/NFI-Up genes.

**Depolarization blocks NFI switch expression via activation of L-type Ca<sup>2+</sup> channels and CaN**

A membrane potential-regulated developmental program was previously identified in which depolarization induced by elevated extracellular [K<sup>+</sup>] inhibited the upregulation of numerous genes expressed in more mature CGNs and stimulated or maintained several genes expressed in immature CGNs (Sato et al., 2005). This voltage-sensitive pattern was similar to that of the NFI switch program, and comparison of the NFI- and KCl-temporal gene cohorts identified 19 commonly regulated temporal genes, including *Gabra6*, *Gabra1*, *Gabrd*, *Camkk2*, *Wnt7a*, *Nptx1*, *Tiam1*, *Slc12a5*, *Kcnk3*, *Dcx*, *Neurod6*, *Tgfb2*, and *Cdh10*. This suggested that at least a portion of the NFI switch program was sensitive to membrane potential in maturing CGNs. qRT-PCR analysis confirmed that numerous Late/NFI-Up genes were inhibited by 25 mM KCl in 6 DIV CGN cultures, while several Early/NFI-Down genes were elevated (Fig. 4A). This included several KCl-regulated genes that were not previously identified by Sato et al. (*Nab2*, *Camk4*, *Egr1*, and *Ect2*). Several KCl-sensitive genes detected by Sato et al. were not regulated by NFI (e.g., *Kcna1*, *Tle1*, and *Prickle1*) (data not shown). Not all temporal mRNAs examined segregated into these two KCl-regulatory/NFI-regulatory patterns; for example, transcripts for the Early/NFI-Down gene *Ect2* were repressed by depolarization, and the Late/NFI-Up gene *Gabrd* was relatively insensitive to 25 mM KCl (Fig. 4A) as previously reported (Sato et al., 2005).

A similar switch pattern was observed in CGNs treated with KA (Fig. 4B), which also depolarizes CGNs, raises intracellular [Ca<sup>2+</sup>] (Courtney et al., 1990), and inhibits maturation-dependent gene expression (Iijima et al., 2009) via activation of AMPA/kainate receptors. Thus, the NFI switch program is more generally sensitive to membrane depolarization-dependent mechanisms.

Elevated KCl increases steady-state Ca<sup>2+</sup> levels in mouse CGNs via L-type voltage-gated calcium channels (VGCCs) (Sato et al., 2005). Nimodipine, a specific inhibitor of L-type VGCCs, derepressed Late-Up gene expression in CGNs cultured in 25 mM KCl (Fig. 4A). In contrast, only one Early/NFI-Down gene, *Dcx*, showed some reversal of the stimulatory effects of depolarization in 6 DIV CGNs, indicating the involvement of additional



**Figure 5.** NFI temporal occupancy is suppressed by depolarization in a CaN-dependent manner. CGNs were cultured in 5 mM or 25 mM KCl  $\pm$  FK506 until 6 DIV and then assayed for NFI occupancy of the indicated Late/NFI-Up genes using ChIP qPCR. Negative control antibody signals were subtracted from ChIP/qPCR values in each case. KCl (25 mM) blocked the temporal increase in NFI binding, and CaN antagonism using FK506 reversed this inhibitory effect; 5, 5 mM KCl; 25, 25 mM KCl; FK, FK506; 25 versus 5:  $^{**}p < 0.01$ ;  $^{***}p < 0.001$ ; (25 + FK) versus 25:  $^{\#}p < 0.05$ ;  $^{\#\#}p < 0.01$ ;  $^{\#\#\#}p < 0.001$ . Data are representative of three independent experiments.

depolarization-sensitive mechanisms in the regulation of this gene subset. This was not further investigated here.

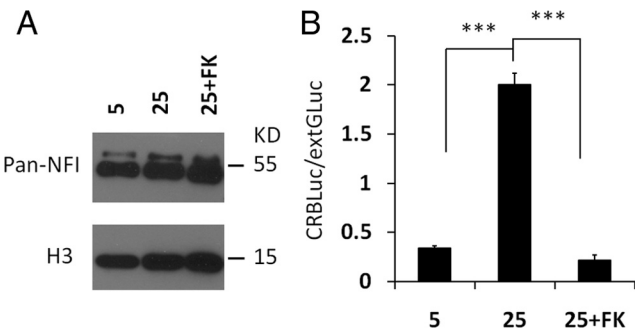
CaN activation has been implicated in depolarization-dependent regulation of numerous genes in CGNs (Guerini et al., 2000; Ives et al., 2002; Kramer et al., 2003; Sato et al., 2005; Iijima et al., 2009). To address its role in the NFI switch, we first confirmed that CaN-dependent dephosphorylation activity was stimulated ( $\sim$ 3-fold) in 6 DIV CGNs cultured in 25 mM KCl and that this activation was blocked by the CaN inhibitor FK506 (Fig. 4C). FK506 reversed inhibition by 25 mM KCl or KA for all Late-Up genes examined (Fig. 4A,B). Similar results were obtained with the CaN antagonist cyclosporine A (data not shown). In contrast to nimodipine, FK506 significantly suppressed depolarization-induced elevation of the Early-Up genes *Dcx*, *Neurod6*, and *Tgfb2*, consistent with earlier findings (Sato et al., 2005).

#### Membrane depolarization and CaN activation prevent NFI temporal binding

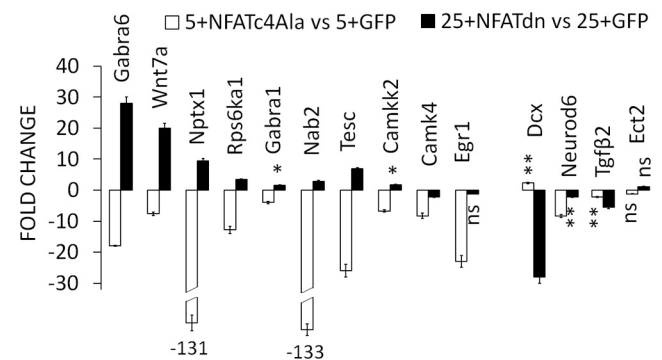
Since delayed occupancy is a central feature of the NFI switch program, we asked whether NFI binding to Late/NFI-Up genes was sensitive to  $K^+$  depolarization. NFI association with chromatin was repressed in 6 DIV CGNs cultured in 25 mM KCl for all six genes tested (*Gabra6*, *Wnt7a*, *Nptx1*, *Tesc*, *Nab2*, *Gabra1*) (Fig. 5). Further, these inhibitory effects were reversed by the CaN inhibitor FK506 (Fig. 5). Thus, temporal NFI occupancy was a primary target of depolarization-induced suppression of Late/Up genes mediated by CaN, including the GABA<sub>A</sub> switch genes *Gabra6* and *Gabra1*.

#### NFAT activation represses the NFI temporal switch

Previous studies found no evidence for differential phosphorylation of NFI proteins in maturing CGNs (Wang et al., 2011). Consistent with this, 25 mM KCl ( $\pm$ FK506) did not alter the amounts of nuclear NFI proteins or their electrophoretic mobilities in maturing CGN cultures (Fig. 6A). This result indicated that CaN activation likely modified other proteins that regulate the NFI switch. NFATc proteins are transcription factors that are regulated via CaN-dependent nuclear translocation (Nguyen and Di Giovanni, 2008), and NFATc4 nuclear localization is stimulated in depolarized neurons, including CGNs (Graef et al., 1999; Benedetto et al., 2005). Using a lentiviral NFAT reporter, we confirmed that depolarization of CGNs stimulated NFAT transcrip-



**Figure 6.** Depolarizing conditions have no effect on NFI proteins but stimulate NFAT transcriptional activity via CaN. **A**, Western blot of NFI proteins in 6 DIV CGNs cultured in 5 mM KCl (5) or in 25 mM KCl (25)  $\pm$  the CaN inhibitor FK506. Histone H3 (H3) served as a loading control. **B**, CGNs were transduced with NFAT dual-reporter lentivirus on 0 DIV and cultured as in **A** to 6 DIV. NFAT transcriptional activity was then determined in cell extracts by assaying luciferase activity.  $^{***}p < 0.001$ . Data are representative of either two (**A**) or three (**B**) independent experiments.

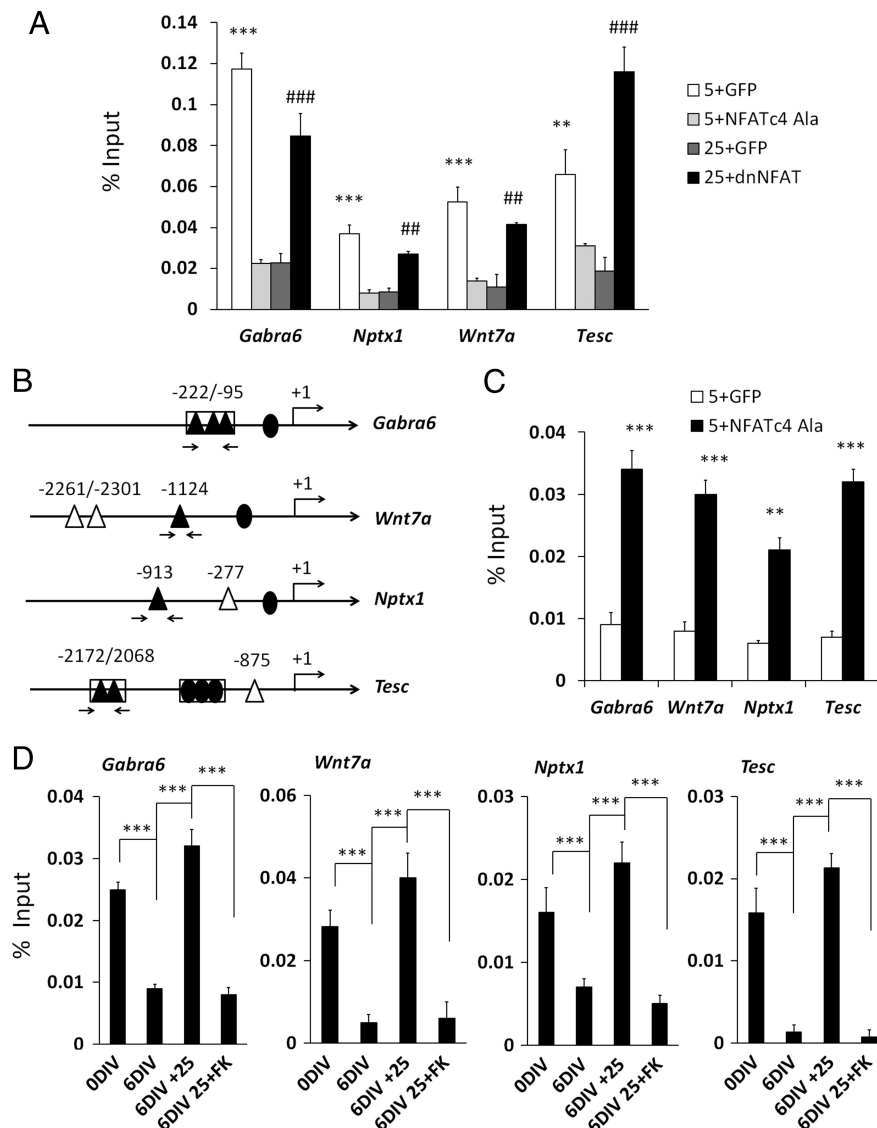


**Figure 7.** NFAT proteins regulate NFI switch gene expression. CGNs were transduced on 0 DIV with GFP or NFATc4-Ala lentiviruses and cultured in 5 mM KCl medium (5), or were transduced with GFP- or dnNFAT-expressing lentiviruses and cultured in depolarizing (25 mM KCl-containing) medium (25). Cells were harvested on 6 DIV and assayed by qRT-PCR. Late/NFI-Up genes are shown on the left, Early/NFI-Down genes on the right.  $p < 0.001$  except  $^*p < 0.05$ ,  $^{**}p < 0.01$ ; ns, no significant difference. Data are representative of three independent experiments.

tional activity (Fig. 6B). We then examined the effects of blocking Ca<sup>2+</sup>/CaN-dependent NFATc nuclear translocation by expressing a dominant-negative form of NFATc4 (dnNFAT) (Chow et al., 1999). DnNFAT significantly reversed depolarization-dependent inhibition of nearly all Late-Up genes tested in CGNs cultured in 25 mM KCl (Fig. 7). Similarly, dnNFAT prevented upregulation of Early-Down genes by 25 mM KCl with the exception of the noncanonical *Ect2* gene, which was unaffected (Fig. 7). Thus, suppression of the NFI switch program by depolarization was largely dependent on activation of endogenous NFAT proteins.

Since nuclear translocation of NFATc4 is stimulated in depolarized neurons, we tested the effects of FLAG-tagged NFATc4 (NFATc4-Ala) that constitutively translocates to the nucleus and regulates transcription (Yang et al., 2002). In CGNs maturing in physiological medium (5 mM KCl), NFATc4-Ala strongly repressed all Late-Up genes examined (Fig. 7). Further, the Early-Down gene *Dcx* was upregulated by NFATc4-Ala while *Neurod6* and *Tgfb2* were inhibited (Fig. 7), suggesting direct or indirect repressive effects on the latter genes due to constitutive NFATc activity. Thus, enhanced NFATc4 activity in nondepolarized CGNs reproduced many of the effects of 25 mM KCl on NFI switch program expression.





**Figure 8.** NFAT proteins regulate NFI binding in depolarized CGNs. **A**, CGNs transduced with lentiviruses expressing NFATc4-Ala, dnNFAT, or GFP were cultured in medium containing 5 or 25 mM KCl to 6 DIV. NFI binding was determined by ChIP qPCR. (5 + NFATc4-Ala) versus (5 + GFP); \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ ; (25 + dnNFAT) versus (25 + GFP); ## $p < 0.01$ ; ### $p < 0.001$ . **B**, NFAT binding sites in the 5'-flanking regions of four Late/NFI-Up gene promoters. Open triangles, weak NFAT binding sites; filled triangles, strong NFAT binding sites; filled ovals, regions of NFI temporal occupancy. Positions of ChIP qPCR primers are shown by arrows. **C**, ChIP qPCR assay of NFATc4-Ala binding to NFAT binding sites (filled triangles in **B**) using anti-FLAG antibody. **D**, ChIP qPCR assay of endogenous NFATc4 binding in CGN cultures in 5 mM (0, 6 DIV) and 25 mM KCl (6 DIV  $\pm$  FK506). **A**, **C**, **D**, Control antibody ChIP values were subtracted to give specific signal, \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ . Data are representative of two (**A**, **C**) or three (**D**) independent experiments.

**NFAT proteins control NFI occupancy in depolarized CGNs**

Since depolarization inhibits NFI temporal interactions with chromatin, we examined whether NFAT proteins specifically regulated NFI binding of Late/NFI-Up genes. We focused on four genes showing the most robust depression by dnNFAT (*Gabra6*, *Wnt7a*, *Nptx1*, *Tesc*). dnNFAT expression reversed the inhibitory effects of depolarizing medium on NFI occupancy in each case (Fig. 8A). Conversely, constitutively active NFATc4-Ala inhibited NFI binding to these genes in CGNs cultured in nondepolarizing (5 mM KCl) medium (Fig. 8A).

The actions of elevated NFAT activity on NFI occupancy may have been direct or indirect. Sequence analysis identified multiple consensus NFAT binding sites near the regions of NFI occupancy within the four Late-Up gene promoters, suggesting

possible direct NFAT interactions (Fig. 8B). To test this, CGN cultures were transduced with FLAG-tagged NFATc4-Ala and assayed by ChIP on 6 DIV. Elevated FLAG signal was detected within specific NFAT-site regions for all four promoters (Fig. 8C). Similarly, endogenous NFATc4 occupancy of Late/NFI-Up genes was markedly increased in nontransduced 6 DIV CGNs cultured in 25 mM KCl (Fig. 8D). Thus, NFATc4 occupancy was elevated under conditions of reduced NFI occupancy of Late/NFI-Up genes, consistent with a direct inhibitory effect of this factor on the NFI switch program in depolarized CGNs.

**Effect of NFAT proteins on dendritogenesis in depolarized CGNs**

Both disruption of NFI function and depolarization inhibit dendrite maturation by CGNs, and CaN activation is required for membrane potential-dependent inhibition (Wang et al., 2007; Okazawa et al., 2009). We therefore examined whether NFAT proteins also mediated membrane potential actions on dendrite formation. Elevated KCl inhibited dendrite length when compared with normal (5 mM) culture medium, and this effect was substantially reversed by dnNFAT (Fig. 9A). Reciprocally, constitutively active NFATc4-Ala repressed CGN dendrite extension in 5 mM KCl cultures (Fig. 9A). NFAT proteins therefore appear to be an integral part of a voltage-sensitive pathway regulating dendrite formation in maturing CGNs.

**Endogenous CaN and NFAT proteins regulate dendrite formation in nondepolarized CGNs**

Maturation-dependent reduction of CaN activity has been proposed to promote the differentiation of postmigratory CGNs within the developing postnatal IGL (Nakanishi and Okazawa, 2006). However, a direct test of CaN function in CGNs differentiating under normal, nondepolarized circumstances was not previously addressed, to our knowledge. Consistent with this hypothesis, CaN activity declined as CGNs matured in physiological culture medium (Fig. 9B). Further, FK506 treatment increased dendrite extension by immature CGNs in 5 mM KCl cultures (Fig. 9C). Similarly, dnNFAT enhanced dendritic length in CGNs maturing in physiological medium (Fig. 9C). Thus, endogenous CaN and NFAT proteins repress dendrite emergence during onset of differentiation by immature CGNs.

**CaN and NFATc4 regulate switch expression and NFI occupancy during normal CGN differentiation**

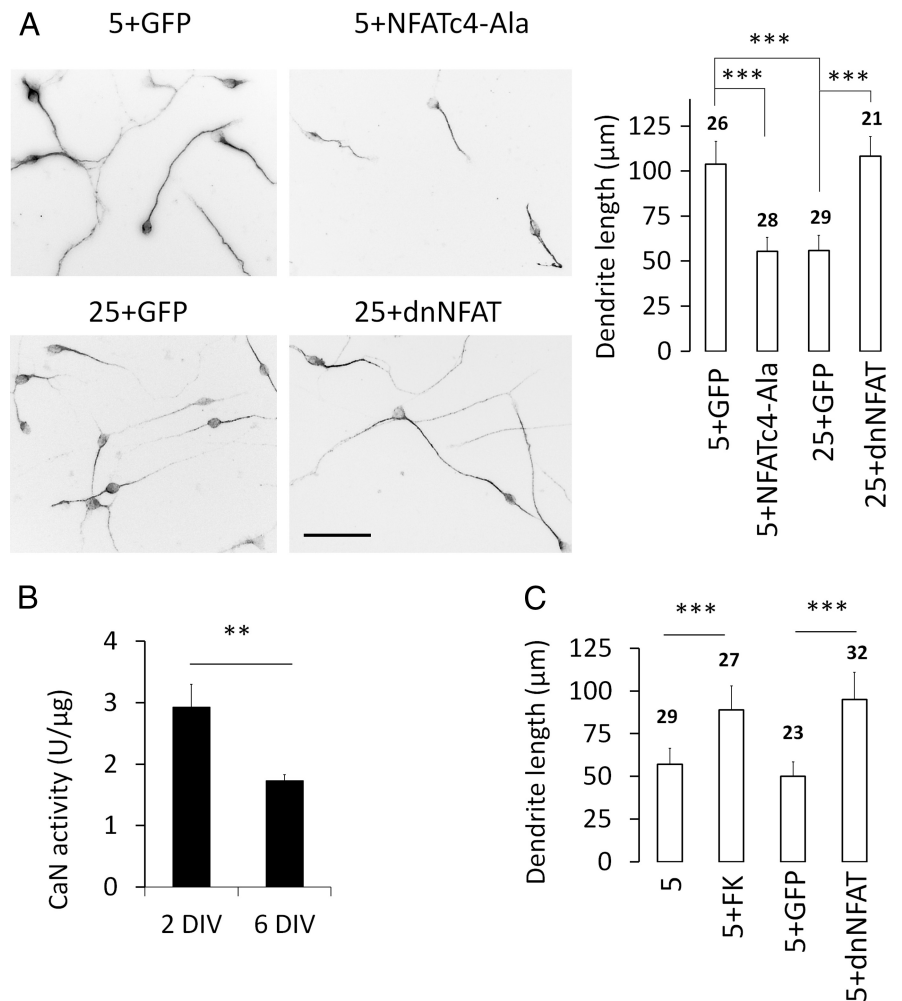
Based on the above findings, we examined whether CaN-dependent mechanisms also regulated the NFI switch program in

CGNs maturing under physiological conditions. Treatment of CGN cultures with FK506 in medium containing 5 mM KCl enhanced Late/NFI-Up gene expression and decreased transcripts for the Early/NFI-Down genes *Dcx* and *Tgfb2* (Fig. 10A). Switch-enhancing effects were observed in both less mature (3 DIV) and more mature (6 DIV) cultures, although they were greater at 3 DIV for several genes, consistent with elevated CaN activity in immature CGNs. These results suggested an inhibitory function for CaN on the NFI switch program during normal differentiation of CGNs.

To examine the role of endogenous NFAT proteins in regulating the NFI switch, we first determined whether endogenous NFATc4 occupied Late/NFI-Up gene promoter sites in CGNs maturing under physiological conditions. Robust NFATc4 binding was detected for the *Gabra6*, *Wnt7a*, *Nptx1*, and *Tesc* promoters in 0 DIV CGNs derived from the EGL/PMZ, as well as in the P7 cerebellum composed mainly of immature CGNs in the IGL (Wang et al., 2011) (Figs. 8D, 10B). Further, NFATc4 occupancy of each site markedly declined in more differentiated, 6 DIV CGN cultures and in the P15 cerebellum (Figs. 8D, 10B), consistent with an inhibitory effect on Late/NFI-Up temporal expression in immature CGNs.

To address NFATc4 function more directly, we examined the impact of NFATc4 deficiency on expression of the NFI temporal program. Cerebella from *Nfatc4* knock-out mice were examined at P7–P9, when expression of the NFI switch program is largely repressed and NFATc4 binding to Late/Up gene promoters is robust, as well as at P15 when NFATc4 occupancy is low and the switch program is highly expressed. Expression of numerous Late/NFI-Up genes was significantly upregulated in *Nfatc4*-null cerebella at each age (Fig. 10C). Further, downregulation of the Early/NFI-Down gene *Dcx* was enhanced during this period. In contrast, *Rbfox3/NeuN*, which is upregulated as CGNs mature (Weyer and Schilling, 2003) and is not regulated by NFI (data not shown), was not altered by *Nfatc4* deficiency (Fig. 10C). Thus, enhanced expression of the NFI switch program in the developing *Nfatc4*-null cerebellum was not due to global changes in maturation-associated gene expression. Western analysis also confirmed that NFI protein levels and mobilities were unchanged in postnatal *Nfatc4* ( $-/-$ ) cerebellum (data not shown).

The pattern of NFATc4 temporal occupancy of Late/NFI-Up genes inversely mirrored that for NFI binding (Figs. 3A, 10B). ChIP analysis revealed that NFI binding to the four Late/NFI-Up promoters increased severalfold in P9 *Nfatc4* ( $-/-$ ) cerebellum relative to WT, and remained significantly elevated at P15 (Fig. 10D). Thus, endogenous NFATc4 inhibits NFI occupancy of Late/NFI-Up genes in the developing postnatal mouse cerebellum.

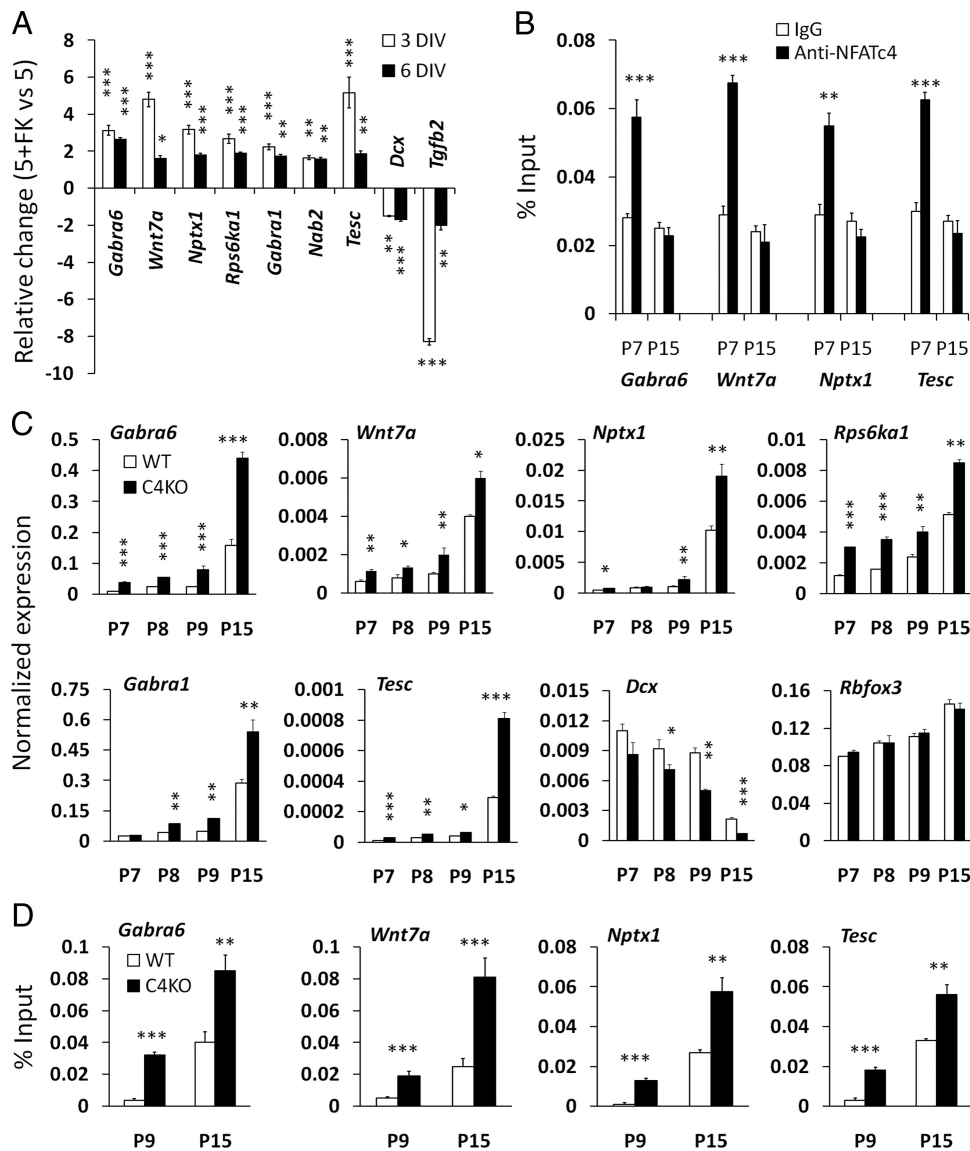


**Figure 9.** Regulation of CGN dendritogenesis by NFAT proteins. **A**, Dendrite length was analyzed on 2 DIV in CGNs transduced with lentiviruses expressing GFP or NFATc4-Ala and cultured in 5 mM KCl medium, and with GFP or dnNFAT viruses and cultured in 25 mM KCl-containing medium. Grayscale images were inverted to provide better visual clarity. Left, Photomicrographs of Map2 immunofluorescence under each condition. Right, Quantification of dendrite length in each case. Scale bar, 50 μm. **B**, CaN activity decreases as CGNs mature. CaN activity was assayed in extracts of immature (2 DIV) and more mature (6 DIV) CGN cultures.  $**p < 0.01$ . **C**, Dendrite lengths for CGNs cultured to 1.5 DIV in standard (5 mM KCl) medium with and without FK506 or transduced with either GFP or dnNFAT viruses. **A**, **C**, The number of cells analyzed under each condition is shown above the histograms.  $***p < 0.001$ . For each part, data are representative of three independent experiments.

## Discussion

The GABA<sub>A</sub> receptor switch and formation of inhibitory synapses between CGNs and GABAergic Golgi type II neurons are critical for cerebellar information flow (Hamann et al., 2002), and their completion coincides with eye opening and enhanced motor activity in rodents (Tia et al., 1996). Thus, the timing of these events is important for developmental onset of sensorimotor function. The present studies reveal a central role for NFI in regulating a late developmental switch program in CGNs that becomes active during a period (P7–P15) dominated by dendritic and synaptic maturation within the IGL. In this process, genes required for mature GABA<sub>A</sub> receptor function and dendrite and synapse formation are upregulated and “immature” genes are downregulated. Delayed NFI occupancy of late-expressed genes is a key feature of this program, coinciding with upregulation of Late/NFI-Up gene expression.

Membrane depolarization is frequently linked with the promotion of neuronal maturation. For example, depolarizing actions of GABA on immature neurons are thought to play a key



**Figure 10.** Regulation of the NFI switch by CaN and NFATc4 in CGNs differentiating in culture and *in vivo*. **A**, Differential expression of Late/NFI-Up genes in standard (5 mM KCl) culture medium with and without FK506. CGN cultures were assayed at 3 (open bars) and 6 (filled bars) DIV. Positive values indicate upregulation and negative values indicate downregulation. **B**, NFATc4 temporal occupancy of Late/NFI-Up genes declines in the developing cerebellum. ChIP qPCR analysis of NFAT binding sites (Fig. 8B) was performed for P7 and P15 mouse cerebellum. **C**, Differential expression of NFI switch genes determined by qRT-PCR in *Nfatc4*-null (C4KO) and WT mouse cerebellum. *Rbfox3* (NeuN) served as a non-NFI regulated negative control. **D**, NFI occupancy was determined by ChIP qPCR in *Nfatc4*-deficient (C4KO) and WT cerebellum at P9 and P15. Negative control antibody values were subtracted to show specific binding. No significant differences except \**p* < 0.05; \*\**p* < 0.01, \*\*\**p* < 0.001. Data are representative of three (**A**, **C**) or two (**B**, **D**) independent experiments.

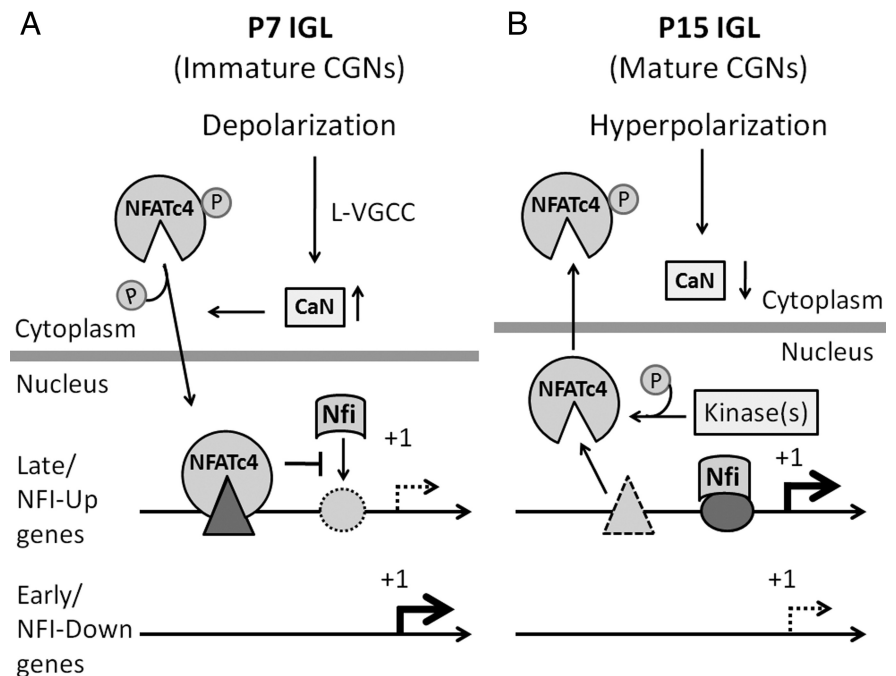
role in corticogenesis and neuronal differentiation (Wang and Kriegstein, 2009; Kilb et al., 2011). However, depolarizing conditions can also block differentiation of neurons, particularly late events linked to dendritic morphology and synaptic development (Sohya et al., 2007; Okazawa et al., 2009). Inhibition of mouse CGN maturation by depolarization-induced CaN activation has been well documented (Guerini et al., 2000; Ives et al., 2002; Kramer et al., 2003; Sato et al., 2005; Iijima et al., 2009; Okazawa et al., 2009). Our studies reveal that the NFI switch program and NFI temporal occupancy are critical targets linking depolarization and CaN activation via L-type VGCCs to inhibition of dendrite formation in CGNs, with NFATc4 being an important intermediary regulator (Fig. 11). Further, these voltage-sensitive mechanisms regulate the NFI switch program and dendrite formation in CGNs differentiating under normal, nondepolarizing conditions.

Our present findings suggest a model in which CaN/NFATc4 inhibit the NFI switch program during development *in vivo*, delaying and modulating the magnitude of a significant portion of its expression in immature postmigratory CGNs in the IGL (Fig. 11). Declining CaN activity and NFATc4 occupancy as CGNs differentiate leads to derepression and is permissive for activation of numerous late-maturation genes associated with dendrite formation, as well as suppression of Early/NFI-Down genes. Downregulation of Early/NFI-Down genes is apparently indirect via undefined repressive factors or signaling pathways that are stimulated by NFI *trans*-activation. Developmental changes in resting membrane potential, which becomes more hyperpolarized as CGNs mature (Rossi et al., 1998; Cathala et al., 2003; Okazawa et al., 2009), appear to regulate late CGN maturation and dendrite formation via Ca<sup>2+</sup>-dependent regulation of CaN (Nakanishi and Okazawa, 2006; Okazawa et al., 2009). Extrinsic and intrinsic

mechanisms that promote hyperpolarization of maturing CGNs may therefore be important upstream drivers of NFI temporal occupancy and the NFI switch within the IGL via CaN/NFATc4 (Fig. 11). An interesting feature of the NFI temporal program is the regulation of several ion channel, transporter and receptor genes that control membrane potential and excitability, including *Slc12a5/Kcc2*, *Kcnk3/Task1*, *Kcnab1/K<sub>v</sub>b1.3*, and GABA<sub>A</sub> receptor subunits. This raises the possibility that NFI temporal programming and CGN excitability may reinforce each other during development, forming a self-amplifying positive feedback loop. Similar mechanisms also may operate in other neurons that undergo developmental membrane hyperpolarization (Ramo and McCormick, 1994; Zhou and Hablitz, 1996; Tyzio et al., 2003).

The timing of neurogenesis has been linked to the formation of discrete patterns of synaptic circuitry for different neuronal subtypes (Tripodi and Arber, 2012). Thus, molecular mechanisms initiated during neuronal birth appear to contribute significantly to the timing of synaptogenesis and circuit formation. In the context of CGN development, postnatal CGNP transplantation experiments showed that maturation-dependent expression of the NFI switch gene *Gabra6* was independent of *in vivo* developmental and regional cues (Bahn et al., 1999). Similarly, *Gabra6* gene expression is maintained in CGNs of HNF3 $\beta$  transgenic mice that fail to properly migrate to the IGL (Zhou et al., 2001). These and other findings suggested that intrinsic timing mechanisms initiated during CGNP cell cycle exit trigger dendritogenesis-associated gene expression in maturing CGNs (Mellor et al., 1998; Bahn et al., 1999). NFI temporal occupancy and its regulation by developmental changes in CaN activity and NFATc4 binding may therefore be part of an internal timing mechanism contributing to dendritic development and its associated gene expression in CGNs. Modulation of these timing mechanisms by environmental factors also may be important for appropriate magnitude and temporality of NFI switch gene expression.

NFI proteins function at multiple stages of CGN development (Wang et al., 2007), and a key question is how they regulate such a diversity of events occurring over varying time frames. The present studies provide insight into their regulation of gene expression specifically required for postmigratory CGN maturation via delayed promoter occupancy. Since NFI proteins and their nuclear DNA binding activity are not temporally upregulated in maturing CGNs (Wang et al., 2011), additional transcriptional and chromatin-related factors presumably contribute to the specificity and timing of their binding and regulation of Late/NFI-Up genes. NFATc4 appears to be an important direct regulator of these events: it binds to numerous Late/NFI-Up gene promoters in immature CGNs, and it is required to prevent premature and excessive NFI occupancy in the developing postnatal cerebellum. The *trans*-repressor REST was recently shown to occupy the *Gabra6* promoter and to inhibit its bind-



**Figure 11.** Model for developmental regulation of the NFI switch program in maturing CGNs. **A**, In the early postnatal cerebellum (P7), CaN activity is elevated in postmigratory CGNs within the IGL, which dephosphorylates NFATc4 and promotes its translocation to the nucleus. NFATc4 binding to Late/NFI-Up gene promoters prevents NFI occupancy and target gene activation. CaN/NFATc4 activities also upregulate expression of Early/NFI-Down genes. Induced depolarization of developing CGNs promotes retention of the immature state via stimulation of Ca<sup>2+</sup> entry via L-type VGCCs and persistent activation of CaN and NFATc4. **B**, Later in development (P15), CaN activity decreases as CGNs mature, which in conjunction with kinase activity reduces NFATc4 occupancy. This is permissive for NFI binding, resulting in increased occupancy of Late/NFI-Up gene promoters and enhanced expression. Reduced CaN/NFATc4 activity also downregulates Early/NFI-Down genes presumably via indirect repressive mechanisms. A shift in resting membrane potential from less to more hyperpolarized in maturing CGNs promotes temporal downregulation of the CaN/NFATc4 pathway and activates NFI temporal binding and the NFI switch during cerebellar development.

ing by NFI in CGNPs and premigratory CGNs derived from the EGL/PMZ (Wang et al., 2011). REST occupancy is depleted in the P7 cerebellum when NFI binding remains low, indicating additional mechanisms function to prevent premature NFI binding in CGNs within the immature IGL. Our findings implicate NFATc4 in the continued repression of NFI occupancy of the *Gabra6* gene in immature postmigratory CGNs in the early postnatal (e.g., P7) IGL following REST dismissal. NFATc4 also limits the magnitude of NFI occupancy of multiple Late/NFI-Up promoters as well as switch program expression in more mature CGNs at P15 (Fig. 10C,D). NFATc4 also may repress Late/NFI-Up genes (and possibly enhance Early/NFI-Down gene expression) via additional, NFI-independent mechanisms.

NFATc4 was previously implicated in neuronal development and function, including axonogenesis and survival, neurotrophin signaling, and memory formation in mice (Graef et al., 2003; Nguyen et al., 2009; Vashishta et al., 2009; Quadrato et al., 2012). Our studies define a novel role for NFATc4 in late CGN development via its temporally downregulated chromatin occupancy. They also identify numerous NFATc4 neuronal targets involved in dendrite/synapse formation, implicating this factor as an important direct regulator of late-maturation gene expression in CGNs.

*Nfia* knock-out mice exhibit severe neurological disruptions, including tremor and diminished cerebellar foliation consistent with altered cerebellar function (das Neves et al., 1999; Wang et al., 2007). Dendrite and synapse formation within the IGL are also altered in *Nfia* (−/−) mice (Wang et al., 2007, 2010), and as

shown here NFIA occupancy of Late/NFI-Up genes is temporally regulated. Further, development of both CGNs and cortical pyramidal neurons is delayed in *Nfia*-null mice (Wang et al., 2007; Piper et al., 2010), consistent with a more general temporal function for this protein in maturing neurons. It is noteworthy that patients with NFIA haploinsufficiency exhibit developmental delays in motor skills and language (Lu et al., 2007; Shaikh et al., 2011), both of which involve cerebellar function (Strick et al., 2009). NFIA and other NFI family members may therefore control important timing mechanisms in CGNs that are involved in developmental delay in both humans and mice.

Formation of distinct patterns of neuronal connectivity has been linked to specific temporal schedules of synaptogenesis (Deguchi et al., 2011; Tripodi and Arber, 2012). Thus, the timing of presynaptic and postsynaptic maturation is a key developmental step in establishing the complexity of neural circuitry that characterizes CNS structure and function. This requires appropriate temporal regulation of gene expression involved in dendritic and synaptic maturation, and disrupted temporal expression can dramatically alter synaptic circuitry (Hippenmeyer et al., 2005; Petrovic and Hummel, 2008). Altered expression of temporally regulated genes during critical time windows affecting synaptic maturation has been proposed to underlie several neurodevelopmental disorders (Meredith et al., 2012). The molecular mechanisms that control these temporal synaptic events are largely unknown. The present studies provide insight into the importance of NFI proteins in the onset of dendrite-related gene expression via their temporally regulated occupancy of synapse-related target genes. An interesting possibility is that altered NFI switch programming and NFI temporal occupancy contribute to disrupted timing of dendrite and synapse formation associated with one or more neurodevelopmental disorders.

## References

- Bahn S, Wisden W, Dunnett SB, Svendsen C (1999) The intrinsic specification of gamma-aminobutyric acid type A receptor alpha6 subunit gene expression in cerebellar granule cells. *Eur J Neurosci* 11:2194–2198. [CrossRef Medline](#)
- Benedito AB, Lehtinen M, Massol R, Lopes UG, Kirchhausen T, Rao A, Bonni A (2005) The transcription factor NFAT3 mediates neuronal survival. *J Biol Chem* 280:2818–2825. [Medline](#)
- Cathala L, Brickley S, Cull-Candy S, Farrant M (2003) Maturation of EPSCs and intrinsic membrane properties enhances precision at a cerebellar synapse. *J Neurosci* 23:6074–6085. [Medline](#)
- Chow CW, Rincón M, Davis RJ (1999) Requirement for transcription factor NFAT in interleukin-2 expression. *Mol Cell Biol* 19:2300–2307. [Medline](#)
- Courtney MJ, Lambert JJ, Nicholls DG (1990) The interactions between plasma membrane depolarization and glutamate receptor activation in the regulation of cytoplasmic free calcium in cultured cerebellar granule cells. *J Neurosci* 10:3873–3879. [Medline](#)
- D'Angelo E, Mazzarello P, Prestori F, Mapelli J, Solinas S, Lombardo P, Cesana E, Gandolfi D, Congi L (2011) The cerebellar network: from structure to function and dynamics. *Brain Res Rev* 66:5–15. [CrossRef Medline](#)
- das Neves L, Duchala CS, Tolentino-Silva F, Haxhiu MA, Colmenares C, Macklin WB, Campbell CE, Butz KG, Gronostajski RM, Godinho F (1999) Disruption of the murine nuclear factor I-A gene (*Nfia*) results in perinatal lethality, hydrocephalus, and agenesis of the corpus callosum. *Proc Natl Acad Sci U S A* 96:11946–11951. [CrossRef Medline](#)
- de la Torre-Ubieta L, Gaudillière B, Yang Y, Ikeuchi Y, Yamada T, DiBacco S, Stegmüller J, Schüller U, Salih DA, Rowitch D, Brunet A, Bonni A (2010) A FOXO-Pak1 transcriptional pathway controls neuronal polarity. *Genes Dev* 24:799–813. [CrossRef Medline](#)
- Deguchi Y, Donato F, Galimberti I, Cabuy E, Caroni P (2011) Temporally matched subpopulations of selectively interconnected principal neurons in the hippocampus. *Nat Neurosci* 14:495–504. [CrossRef Medline](#)
- Di Cristo G, Chattopadhyaya B, Kuhlman SJ, Fu Y, Bélanger MC, Wu CZ, Rutishauser U, Maffei L, Huang ZJ (2007) Activity-dependent PSA expression regulates inhibitory maturation and onset of critical period plasticity. *Nat Neurosci* 10:1569–1577. [CrossRef Medline](#)
- Furuichi T, Shiraishi-Yamaguchi Y, Sato A, Sadakata T, Huang J, Shinoda Y, Hayashi K, Mishima Y, Tomomura M, Nishibe H, Yoshikawa F (2011) Systematizing and cloning of genes involved in the cerebellar cortex circuit development. *Neurochem Res* 36:1241–1252. [CrossRef Medline](#)
- Geschwind DH, Levitt P (2007) Autism spectrum disorders: developmental disconnection syndromes. *Curr Opin Neurobiol* 17:103–111. [CrossRef Medline](#)
- Goldowitz D, Hamre K (1998) The cells and molecules that make a cerebellum. *Trends Neurosci* 21:375–382. [CrossRef Medline](#)
- Graef IA, Mermelstein PG, Stankunas K, Neilson JR, Deisseroth K, Tsien RW, Crabtree GR (1999) L-type calcium channels and GSK-3 regulate the activity of NF-ATc4 in hippocampal neurons. *Nature* 401:703–708. [CrossRef Medline](#)
- Graef IA, Wang F, Charron F, Chen L, Neilson J, Tessier-Lavigne M, Crabtree GR (2003) Neurotrophins and netrins require calcineurin/NFAT signaling to stimulate outgrowth of embryonic axons. *Cell* 113:657–670. [CrossRef Medline](#)
- Guerini D, Wang X, Li L, Genazzani A, Carafoli E (2000) Calcineurin controls the expression of isoform 4CII of the plasma membrane Ca(2+) pump in neurons. *J Biol Chem* 275:3706–3712. [CrossRef Medline](#)
- Hall AC, Lucas FR, Salinas PC (2000) Axonal remodeling and synaptic differentiation in the cerebellum is regulated by WNT-7a signaling. *Cell* 100:525–535. [CrossRef Medline](#)
- Hamann M, Rossi DJ, Attwell D (2002) Tonic and spillover inhibition of granule cells control information flow through cerebellar cortex. *Neuron* 33:625–633. [CrossRef Medline](#)
- Hippenmeyer S, Vrieseling E, Sigrist M, Portmann T, Laengle C, Ladle DR, Arber S (2005) A developmental switch in the response of DRG neurons to ETS transcription factor signaling. *PLoS Biol* 3:e159. [CrossRef Medline](#)
- Iijima T, Emi K, Yuzaki M (2009) Activity-dependent repression of Cbln1 expression: mechanism for developmental and homeostatic regulation of synapses in the cerebellum. *J Neurosci* 29:5425–5434. [CrossRef Medline](#)
- Ito S, Takeichi M (2009) Dendrites of cerebellar granule cells correctly recognize their target axons for synaptogenesis in vitro. *Proc Natl Acad Sci U S A* 106:12782–12787. [CrossRef Medline](#)
- Ives JH, Drewery DL, Thompson CL (2002) Neuronal activity and its influence on developmentally regulated GABA(A) receptor expression in cultured mouse cerebellar granule cells. *Neuropharmacology* 43:715–725. [CrossRef Medline](#)
- Kilb W, Kirischuk S, Luhmann HJ (2011) Electrical activity patterns and the functional maturation of the neocortex. *Eur J Neurosci* 34:1677–1686. [CrossRef Medline](#)
- Kilpatrick DL, Wang W, Gronostajski R, Litwack ED (2012) Nuclear factor I and cerebellar granule neuron development: an intrinsic-extrinsic interplay. *Cerebellum* 11:41–49. [CrossRef Medline](#)
- Kramer D, Fresu L, Ashby DS, Freeman TC, Genazzani AA (2003) Calcineurin controls the expression of numerous genes in cerebellar granule cells. *Mol Cell Neurosci* 23:325–330. [CrossRef Medline](#)
- Laurie DJ, Wisden W, Seeburg PH (1992) The distribution of thirteen GABAA receptor subunit mRNAs in the rat brain. III. Embryonic and postnatal development. *J Neurosci* 12:4151–4172. [Medline](#)
- Leonardo ED, Hen R (2008) Anxiety as a developmental disorder. *Neuropsychopharmacology* 33:134–140. [CrossRef Medline](#)
- Lu W, Quintero-Rivera F, Fan Y, Alkuraya FS, Donovan DJ, Xi Q, Turbe-Doan A, Li QG, Campbell CG, Shanske AL, Sherr EH, Ahmad A, Peters R, Rilliet B, Parvex P, Bassuk AG, Harris DJ, Ferguson H, Kelly C, Walsh CA, et al. (2007) NFIA haploinsufficiency is associated with a CNS malformation syndrome and urinary tract defects. *PLoS Genet* 3:e80. [CrossRef Medline](#)
- Manzini MC, Ward MS, Zhang Q, Lieberman MD, Mason CA (2006) The stop signal revised: immature cerebellar granule neurons in the external germinal layer arrest pontine mossy fiber growth. *J Neurosci* 26:6040–6051. [CrossRef Medline](#)
- Mellor JR, Merlo D, Jones A, Wisden W, Randall AD (1998) Mouse cerebellar granule cell differentiation: electrical activity regulates the GABAA receptor alpha 6 subunit gene. *J Neurosci* 18:2822–2833. [Medline](#)
- Meredith RM, Dawitz J, Kramvis I (2012) Sensitive time-windows for susceptibility in neurodevelopmental disorders. *Trends Neurosci* 35:335–344. [CrossRef Medline](#)
- Na IK, Markley JC, Tsai JJ, Yim NL, Beattie BJ, Klose AD, Holland AM, Ghosh

- A, Rao UK, Stephan MT, Serganova I, Santos EB, Brentjens RJ, Blasberg RG, Sadelain M, van den Brink MR (2010) Concurrent visualization of trafficking, expansion, and activation of T lymphocytes and T-cell precursors in vivo. *Blood* 116:e18–e25. [CrossRef Medline](#)
- Nakanishi S, Okazawa M (2006) Membrane potential-regulated Ca<sup>2+</sup> signaling in development and maturation of mammalian cerebellar granule cells. *J Physiol* 575:389–395. [CrossRef Medline](#)
- Nguyen T, Di Giovanni S (2008) NFAT signaling in neural development and axon growth. *Int J Dev Neurosci* 26:141–145. [CrossRef Medline](#)
- Nguyen T, Lindner R, Tedeschi A, Forsberg K, Green A, Wuttke A, Gaub P, Di Giovanni S (2009) NFAT-3 is a transcriptional repressor of the growth-associated protein 43 during neuronal maturation. *J Biol Chem* 284:18816–18823. [CrossRef Medline](#)
- Okazawa M, Abe H, Katsukawa M, Iijima K, Kiwada T, Nakanishi S (2009) Role of calcineurin signaling in membrane potential-regulated maturation of cerebellar granule cells. *J Neurosci* 29:2938–2947. [CrossRef Medline](#)
- Petrovic M, Hummel T (2008) Temporal identity in axonal target layer recognition. *Nature* 456:800–803. [CrossRef Medline](#)
- Piper M, Barry G, Hawkins J, Mason S, Lindwall C, Little E, Sarkar A, Smith AG, Moldrich RX, Boyle GM, Tole S, Gronostajski RM, Bailey TL, Richards LJ (2010) NFIA controls telencephalic progenitor cell differentiation through repression of the Notch effector Hes1. *J Neurosci* 30:9127–9139. [Medline](#)
- Piper M, Harris L, Barry G, Heng YH, Plachez C, Gronostajski RM, Richards LJ (2011) Nuclear factor one X regulates the development of multiple cellular populations in the postnatal cerebellum. *J Comp Neurol* 519:3532–3548. [CrossRef Medline](#)
- Puzianowska-Kuznicka M, Shi YB (1996) Nuclear factor I as a potential regulator during postembryonic organ development. *J Biol Chem* 271:6273–6282. [CrossRef Medline](#)
- Quadrato G, Benevento M, Alber S, Jacob C, Floriddia EM, Nguyen T, Elnagar MY, Pedroarena CM, Molkentin JD, Di Giovanni S (2012) Nuclear factor of activated T cells (NFATc4) is required for BDNF-dependent survival of adult-born neurons and spatial memory formation in the hippocampus. *Proc Natl Acad Sci U S A*.
- Raetzman LT, Siegel RE (1999) Immature granule neurons from cerebella of different ages exhibit distinct developmental potentials. *J Neurobiol* 38:559–570. [CrossRef Medline](#)
- Ramoa AS, McCormick DA (1994) Developmental changes in electrophysiological properties of LGNd neurons during reorganization of retinogeniculate connections. *J Neurosci* 14:2089–2097. [Medline](#)
- Rossi DJ, Hamann M (1998) Spillover-mediated transmission at inhibitory synapses promoted by high affinity alpha6 subunit GABA(A) receptors and glomerular geometry. *Neuron* 20:783–795. [CrossRef Medline](#)
- Rossi P, De Filippi G, Armano S, Taglietti V, D'Angelo E (1998) The weaver mutation causes a loss of inward rectifier current regulation in premigratory granule cells of the mouse cerebellum. *J Neurosci* 18:3537–3547. [Medline](#)
- Sato M, Suzuki K, Yamazaki H, Nakanishi S (2005) A pivotal role of calcineurin signaling in development and maturation of postnatal cerebellar granule cells. *Proc Natl Acad Sci U S A* 102:5874–5879. [CrossRef Medline](#)
- Shaikh TH, Haldeman-Englert C, Geiger EA, Ponting CP, Webber C (2011) Genes and biological processes commonly disrupted in rare and heterogeneous developmental delay syndromes. *Hum Mol Genet* 20:880–893. [CrossRef Medline](#)
- Sohya K, Kitamura A, Akaneya Y (2007) Chronic membrane depolarization-induced morphological alteration of developing neurons. *Neuroscience* 145:232–240. [CrossRef Medline](#)
- Strick PL, Dum RP, Fiez JA (2009) Cerebellum and nonmotor function. *Annu Rev Neurosci* 32:413–434. [CrossRef Medline](#)
- Tia S, Wang JF, Kotchabhakdi N, Vicini S (1996) Developmental changes of inhibitory synaptic currents in cerebellar granule neurons: role of GABA(A) receptor alpha 6 subunit. *J Neurosci* 16:3630–3640. [Medline](#)
- Tripodi M, Arber S (2012) Regulation of motor circuit assembly by spatial and temporal mechanisms. *Curr Opin Neurobiol* 22:615–623. [CrossRef Medline](#)
- Tyzio R, Ivanov A, Bernard C, Holmes GL, Ben-Ari Y, Khazipov R (2003) Membrane potential of CA3 hippocampal pyramidal cells during postnatal development. *J Neurophysiol* 90:2964–2972. [CrossRef Medline](#)
- Vashishta A, Habas A, Pruunsild P, Zheng JJ, Timmusk T, Hetman M (2009) Nuclear factor of activated T-cells isoform c4 (NFATc4/NFAT3) as a mediator of antiapoptotic transcription in NMDA receptor-stimulated cortical neurons. *J Neurosci* 29:15331–15340. [CrossRef Medline](#)
- Wang DD, Kriegstein AR (2009) Defining the role of GABA in cortical development. *J Physiol* 587:1873–1879. [CrossRef Medline](#)
- Wang W, Stock RE, Gronostajski RM, Wong YW, Schachner M, Kilpatrick DL (2004) A role for nuclear factor I in the intrinsic control of cerebellar granule neuron gene expression. *J Biol Chem* 279:53491–53497. [CrossRef Medline](#)
- Wang W, Qu Q, Smith FI, Kilpatrick DL (2005) Self-inactivating lentiviruses: versatile vectors for quantitative transduction of cerebellar granule neurons and their progenitors. *J Neurosci Methods* 149:144–153. [CrossRef Medline](#)
- Wang W, Mullikin-Kilpatrick D, Crandall JE, Gronostajski RM, Litwack ED, Kilpatrick DL (2007) Nuclear factor I coordinates multiple phases of cerebellar granule cell development via regulation of cell adhesion molecules. *J Neurosci* 27:6115–6127. [CrossRef Medline](#)
- Wang W, Crandall JE, Litwack ED, Gronostajski RM, Kilpatrick DL (2010) Targets of the nuclear factor I regulon involved in early and late development of postmitotic cerebellar granule neurons. *J Neurosci Res* 88:258–265. [CrossRef Medline](#)
- Wang W, Shin Y, Shi M, Kilpatrick DL (2011) Temporal control of a dendritogenesis-linked gene via REST-dependent regulation of nuclear factor I occupancy. *Mol Biol Cell* 22:868–879. [CrossRef Medline](#)
- Weyer A, Schilling K (2003) Developmental and cell type-specific expression of the neuronal marker NeuN in the murine cerebellum. *J Neurosci Res* 73:400–409. [CrossRef Medline](#)
- Yang TT, Xiong Q, Ensen H, Davis RJ, Chow CW (2002) Phosphorylation of NFATc4 by p38 mitogen-activated protein kinases. *Mol Cell Biol* 22:3892–3904. [CrossRef Medline](#)
- Zheng T, Santi MR, Bovolenta P, Marlier LN, Grayson DR (1993) Developmental expression of the alpha 6 GABAA receptor subunit mRNA occurs only after cerebellar granule cell migration. *Brain Res Dev Brain Res* 75:91–103. [Medline](#)
- Zhou FM, Hablitz JJ (1996) Postnatal development of membrane properties of layer I neurons in rat neocortex. *J Neurosci* 16:1131–1139. [Medline](#)
- Zhou H, Hughes DE, Major ML, Yoo K, Pesold C, Costa RH (2001) Atypical mouse cerebellar development is caused by ectopic expression of the forkhead box transcription factor HNF-3beta. *Gene Expr* 9:217–236. [Medline](#)